

First results from a controlled deep-sea CO₂ perturbation experiment: Evidence for rapid equilibration of the oceanic CO₂ system at depth

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Key Words

Carbon Dioxide
Seawater
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pH

Abstract

We have carried out series of ROV controlled oceanic CO₂ system perturbation experiments off the coast of California at depths down to 1000m to observe reaction rates and pathways with both HCl and HCO₃⁻ addition. The work was done to evaluate possible barriers to carrying out future FOCE (Free Ocean CO₂ Enrichment) experiments to simulate the chemistry of the emerging high CO₂-lower pH ocean. A looped 460 ml flow cell with a pH sensor was used to monitor the time-to-equilibrium for 900μl additions of 0.008N HCl, and for small slugs of HCO₃⁻ enriched sea water. The results were compared to equivalent experiments at the same temperature and 1 atmosphere pressure. In each case the experiments at depth showed significantly faster time-to-equilibrium than those at one atmosphere. These results are consistent with the low partial molal volume of CO₂ in sea water favoring the hydration reaction rate. The results imply, but do not prove, a significant effect of pressure on the rate constants. The relatively rapid equilibration times observed in sea water of 4°C and at 10 MPa indicates that there are no fundamental physical chemistry limits for carrying out small scale free ocean CO₂ enrichment experiments.

Introduction

Background

It is now commonplace to carry out large-scale controlled CO₂ enrichment experiments on land to investigate the impact of increasing atmospheric CO₂ levels on terrestrial ecosystems, but no equivalent experiments have yet been carried out in the ocean. Why is that, is there a compelling need, and what are the conflicts and challenges?

The extraordinary rise in fossil fuel CO₂ levels in the atmosphere has as its partner an equivalent rise in the CO₂ concentration in the ocean. This ocean has long been recognized as “a giant regulator of carbon dioxide [Callendar, 1938], and with the ability to recover the oceanic fossil fuel CO₂ signal above the natural background [Brewer, 1978; Gruber *et al.*, 1996] scientists have been able to track the increasing oceanic CO₂ levels. About 30% of the fossil fuel CO₂ emitted to the atmosphere is quickly transferred by gas exchange to the ocean surface waters [IPCC, 1990, 1995], and the rate of ocean uptake is now ~ 20-25 million metric tons CO₂ per day (2 –2.5 Gt C/yr). Without this vast present and future sink provided by the alkalinity of the ocean the world would have an extraordinary atmospheric CO₂ problem. We explicitly count upon continued oceanic uptake in all extrapolations of the future accumulation of CO₂ in the atmosphere.

Yet these same projections also indicate that an unprecedented change in oceanic chemistry may occur [Haugan and Drange, 1996; Brewer, 1997; Caldeira and Wickett, 2003] that if taken too far may cause environmental harm [Kleypas *et al.*, 1999; Langdon *et al.*, 2000; Riebesell *et al.*, 2000]. What level is “too far”? – and how might we assess this? We have already reduced the pH of ocean surface waters worldwide by >0.1 pH units, and when CO₂ levels reach 600 ppm in mid-century we will have reduced the pH by 0.3. All future energy scenarios have continued fossil fuel usage, and thus a lower pH ocean is inevitable with only the time scale at issue.

On land the climatic effects of elevated CO₂ levels have been treated both as a problem – global warming, sea level rise, but also as a possible benefit in increased plant photosynthesis. It has even been suggested that the increase in terrestrial photosynthesis could greatly offset national contributions to atmospheric CO₂ levels [Fan *et al.*, 1998; DeLucia *et al.*, 1999], although this result has been strongly criticized [Holland and Brown, 1999; Potter and Klooster, 1999; Bolin *et al.*, 1999].

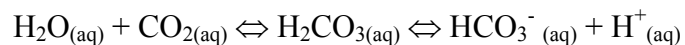
The experimental basis for evaluating land ecosystem responses to elevated CO₂ levels has been greatly aided by the development of Free Air CO₂ Enrichment (FACE) techniques in which a ring of controlled CO₂ diffusers release gas on the upwind side of the apparatus so that downstream sensors receive the signal and provide feedback to control the flow. In this way long-term controlled exposure of a complex, open ecosystem to a set elevated atmospheric CO₂ level is maintained. No such techniques are yet available for the equivalent oceanic experiments.

Oceanic CO₂ Experiments

Large-scale aquarium experiments [Langdon *et al.*, 2000], mesocosm studies [Riebesell *et al.*, 2000; Zondervan *et al.*, 2001], and small-scale studies of the effect of elevated oceanic CO₂ levels on animal reproduction [Shirayama, 1997] have been carried out. Deep-ocean CO₂ sequestration experiments with free release of liquid CO₂ [Brewer *et al.*; 1999, 2002; Barry *et al.*, 2003] have provided valuable information, but tidal vector forcing of the plume of CO₂ rich, low pH water emanating from a central pool results in complex and highly variable exposure of the surrounding biogeochemical system. Carrying out the oceanic equivalent of the land based FACE experiments could alleviate this problem.

There are however substantial technical barriers to doing so. With the new availability of advanced ROV techniques for experimental ocean science we can visualize ways to assemble, control, and return data from, complex geochemical experiments on the ocean floor [Kleinberg *et al.*, 2003; Brewer *et al.*, 2004; Pasteris *et al.*, 2004]. But difficult fundamental chemical challenges also arise; CO₂ released into free air is inert – only physical mixing is required to distribute the signal. Release of CO₂ into sea water initiates a complex series of reactions involving the full aqueous CO₂-H₂CO₃-HCO₃⁻-CO₃⁼ system. And while these species are typically treated as an equilibrium system, the slow hydration rate of the CO₂ molecule (Johnson, 1982; Soli and Byrne, 2002) can produce very significant lag times that can potentially confound an oceanic CO₂ perturbation experiment [Zeebe *et al.*, 1999; Zeebe and Wolf-Gladrow, 2001].

The essential reaction at typical sea water pH is



Where the initial non-ionic reaction is slow with an Arrhenius activation energy of 74 kJ/mol [Magid and Turbeck, 1968].

The effect of pressure on the equilibrium state of chemical species in sea water can readily be calculated with knowledge of the partial molal volumes of the reactants, and this is routinely done for calculating the *in situ* properties of the oceanic CO₂ system. But there is no way to compute a priori the effects of pressure on the reaction rates. That must be determined experimentally.

We report here an investigation of this problem, carried out through a series of small-scale time-to-equilibrium experiments at various ocean depths by injecting both hydrochloric acid, and CO₂ enriched sea water, into a looped flow cell and observing the response time at depth.

Methods

Apparatus

The experiments created here are simple in concept, but difficult to execute. We constructed a flow loop system (Figs. 1 and 2) that permitted sea water at a selected depth to be drawn in to the unit, and circulated through a flow cell equipped with a pH electrode (Seabird Electronics, Inc., SBE 18) and a thermistor for logging temperature. The internal volume of the cell and associated valves and tubing was approximately 460ml. Hydraulic valves, actuated under ROV control, were used to open and close the inlet and outlet ports. Data logging was by laptop computer in the vehicle control room, using MBARI created software. A screen image of the data acquisition system is shown in Fig. 3.

For the acid injection experiments a 1 ml syringe permitted injection of 900 μL of 0.008N HCl in 0.5 M NaCl into the cell at each stroke, much as in a classic alkalinity titration [Bradshaw *et al.*, 1981]. A small capillary loop, open to the ocean, served to accommodate the volume increase.

For the CO₂ enriched water injections an inverted box, crafted from cutting a laboratory polyacrylic carboy, was attached to the frame to contain approximately 1L liquid CO₂ delivered at depth following techniques described earlier [Brewer *et al.*, 1999]. Sea water trapped in close contact with the liquid was strongly enriched in dissolved inorganic carbon (DIC), primarily in the form of HCO₃⁻ ion, and a slug of this water could be drawn into the cell by using the vehicle robotic arm to place a wand, with inlet tube attached, in the pool of CO₂ rich water. The hydraulic valves were then rapidly opened and closed with the re-circulating pump running, so that a small slug (of indeterminate volume) was introduced into the circulating flow.

Field work

The experiments described here took place on January 27 and 28, 2004 in Monterey Bay, California, using the RV Point Lobos, and the ROV Ventana as experimental platforms for equipment deployment and control. Bad weather on January 27 curtailed activities, and work on that day served mainly as a test to assure that the system would yield reliable data, and to make small adjustments to the point of delivery of the acid so that the low pH water at the capillary tip passed once through the circulation loop to become well mixed before contacting the pH electrode surface.

The procedure followed on January 28, 2004 was to dive the vehicle to 1000m depth, and carry out a set of three acid injections, followed by a series of three injections of highly CO₂ enriched water. Both single strokes of the syringe, and double strokes, were used so as to examine the relaxation times at differing ΔpH. The cell configuration remained the same for all dives, and each experiment was allowed to run for 5 minutes. The vehicle was then brought to shallower depth, and the exercise repeated at 500m, and at 100m depth.

Results

The raw sensor (electrode + preamplifier) output voltage obtained was first converted to pH by using the equation

$$\text{pH} = 7 + (V_{\text{out}} - \text{offset}) / (K \times T \times \text{slope})$$

where, V_{out} is the pH electrode output voltage, $K = (R/F) \times \ln 10 = 1.98416 \times 10^{-4} \text{ V K}^{-1}$, R is the gas constant ($8.31434 \text{ K}^{-1} \text{ mol}^{-1}$), F is the Faraday constant ($9.6486 \times 10^4 \text{ C mol}^{-1}$), and T is temperature in Kelvin. In the following discussion, all times are given in UTC as recorded by the ship/ROV experimental system.

The first experiment at 1000 m depth, $T = \sim 4^\circ\text{C}$, $S = 34.517$, was initiated at 18:02h with injection of single 900 μL stroke of HCl. These single stroke acid injections were carried out three times, and then a double stroke was also done three times under same conditions. Our on-site review of the data revealed small inconsistencies in equilibration times, and this was traced to inadequate flushing of the dead space between the valves between experiments. Experimental procedure was therefore altered to adjust for this at other depths (Figure 4).

Injection of CO₂ enriched water at 1000 m depth was initiated at 19:19h, after the HCl injection experiments were complete, and repeated 4 times. These indicate that the sharp peak of pH at the time of injection of CO₂ enriched water is associated with the opening and closing of the hydraulic valve operation. We observed a system decrease about 0.2 pH units immediately, followed by relaxation to a new equilibrium condition; the kinetics of this process includes both chemical equilibration and water mixing. We continued similar HCl injection and CO₂ enriched seawater injection experiments at 500m depth, $T = \sim 6^\circ\text{C}$, $S = 34.301$, and 100m depth, $T = \sim 10^\circ\text{C}$, $S = 33.683$. The plots of pH versus time at 500m depth are shown in Figs. 4 and 5 for a series of HCl injection, and CO₂ enriched seawater injection, runs respectively.

The relaxation to equilibrium was approached from different directions in these series once the initial large drop in pH occurred. Although both injection series result in lowered pH they differ in important details. For acid injection we change pH and alkalinity, but not the mass of CO₂. For the CO₂ enriched water addition we do not change alkalinity. For the acid addition the volume added is small and tightly specified. For the CO₂ enrichments the volume is larger, and not precisely known.

Both the forward and back reactions are proceeding simultaneously in each scheme. For the acid injection the system first overshoots, then trends to lower pH with time. This is consistent with formation of an intense cloud of CO₂ at the point of injection, followed by slower hydration of the excess of CO₂ molecules over the equilibrium value to form the equilibrium quantity of HCO₃⁻.

For the HCO₃⁻ enriched sea water injections the net result is a rapid decrease from injection of low pH water, followed by a slower increase in pH as the system equilibrates. This is consistent with removal of a temporary excess of HCO₃⁻ over the equilibrium value.

We have fit the data obtained to an exponential relationship. For the HCl injection experiments, the relaxation time (f) of a perturbation in pH is given by

$$f = y_0 + a * (\exp(-b * t))$$

For the CO₂ enriched sea water experiments we use the function

$$f = y_0 + a * (1 - \exp(-b * t))$$

where in both cases t is the time in seconds, and y_0 , a , b are the determined coefficients.

The results of this analysis are shown in Figures 6 and 7 where representative equilibration curves in pH with time for an experiment at 500m depth are shown.

In each case the relaxation time to a new equilibrium value was far shorter than that experienced in laboratory studies at one atmosphere at the same temperature. The results of the half-lives obtained from these *in situ* experiments, together with results from laboratory studies for comparison, are shown in Table 1. The coefficients derived are shown in Table 2.

In each case the time to equilibrium observed was faster for the acid injection experiments than for the introduction of HCO₃⁻ enriched sea water (Table 1). We hypothesize that this is due to the relative volume changes as equilibrium is approached. For an excess of free CO₂ molecules converting to HCO₃⁻ the ΔV is negative, and thus higher pressure favors the reaction. For an excess of HCO₃⁻ the sign is reversed, and equilibrium is reached more slowly. Both these effects are small compared to the overall increase in rates observed.

While the exact numerical values derived here are somewhat apparatus dependent in that they include a mixing term there is one clear trend. In similar laboratory experiments of HCl injection to surface seawater at one atmosphere (Table 2), the trend of the relaxation times observed, as is widely known, is to become slower with decreasing temperature. In the *in situ* experiments in each case the relaxation time was faster at lower temperature. These experiments suggest that the high pressures in the deep sea have a strong impact on the equilibration time, overcoming the effect of lower temperature.

Discussion

The problems of very long times-to-equilibrium for the classic TCO₂-alkalinity acid titration of sea water [Dyrssen and Sillen, 1967; Bradshaw *et al.*, 1981] for small perturbations near the initial pH are familiar to most ocean chemists. It is in large part for this reason that most laboratory titrations are run at 25°C. At low temperatures the time scales are frustratingly long and thus the determination of the apparent stability constants of the CO₂ system in sea water at low temperature is notoriously difficult. Thus it was with some surprise that we first observed the extraordinarily speed with which equilibrium was reached at 4°C, but at 10 MPa pressure.

Surprisingly, the reaction pathway for this system is not well known. Johnson (1982) used transition state theory, in which the reactants are in equilibrium with an activated complex that dissociates to form the products, to correct his results on the rate constants for sea water, but did not specify the complex. The reaction involves breaking of one of the CO₂ double bonds and is also accompanied by a bending of the CO₂ molecule. The reaction is approximately athermic.

An answer was provided by Jonsson *et al.* [1977,1978], who carried out *ab initio* molecular orbital calculations of the reaction $\text{H}_2\text{O} + \text{CO}_2 \rightarrow \text{H}_2\text{CO}_3$, and the reaction $\text{OH}^- + \text{CO}_2 \rightarrow \text{HCO}_3^-$, and identified a possible transition state (Fig. 8). They found no barrier to reaction in the gaseous state, and suggested that the slow reaction in solution must therefore be associated with solvation effects. Specifically they suggested that the transition state identified from the reaction with OH⁻ in the gas phase should also be valid for the reaction $\text{H}_2\text{O}_{(\text{aq})} + \text{CO}_{2(\text{aq})} \leftrightarrow \text{H}_2\text{CO}_{3(\text{aq})}$. We therefore hypothesize that the increase in reaction rate with pressure is associated with changes in the solvation sphere surrounding the dangling proton in the model of Jonsson *et al.* (1978).

The exact value of the enhancement in rate due to pressure effects cannot be measured accurately here. The apparatus we have used is designed for *in situ* work, and cannot be easily adapted to laboratory studies. The precise response time of the system is dependent on experimental variables and is a diagnostic, not a fundamental, number. In fieldwork at a single site we cannot vary pressure and temperature independently. Nonetheless the rapidity of the observed time to equilibrium at depth provides convincing evidence of the feasibility of carrying out controlled deep ocean CO₂ perturbation experiments that may help uncover the emerging effects of steadily increasing ocean acidification on marine biogeochemical and ecological systems. The surprising increase in reaction rate we have uncovered may have significance for the respiration of marine animals at depth.

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References

Barry, J.P., B.A. Seibel, J.C. Drazen, T.M.N. Tamburri, K.R. Buck, C. Lovera, L. Kuhnz, E.T. Peltzer, K. Osborn, P.J. Whaling, P. Walz, and P.G. Brewer, Deep-Sea field experiments on the biological impacts of direct deep-sea CO₂ injection. In: *“Proceedings of the Second Annual Conference on Carbon Sequestration”* U.S. Dept. of Energy, 2003.

Bolin, B., J. Canadell, B. Moore III, I. Noble, and W. Steffen, Effect on the biosphere of elevated atmospheric CO₂, *Science*, 285, 1851-1852, 1999.

Bradshaw, A.L., P.G. Brewer, D.K. Shafer, and R.T. Williams, Measurements of total carbon dioxide and alkalinity by potentiometric titration in the Geosecs program, *Earth Planet. Sci. Lett.*, 55, 99-115, 1981.

Brewer, P. G., Direct observation of the oceanic CO₂ increase. *Geophys. Res. Lett.*, 5, 997-1000, 1978.

Brewer, P.G., Ocean chemistry of the fossil fuel CO₂ signal: the haline signature of “Business as Usual”, *Geophys. Res. Lett.*, 24, 1367-1369, 1997.

Brewer, P.G., G. Friederich, E.T. Peltzer, and F.M. Orr Jr., Direct experiments on the ocean disposal of fossil fuel CO₂, *Science*, 284, 943-945, 1999.

Brewer, P.G., E. T. Peltzer, G. Friederich, and G. Rehder, Experimental determination of the fate of rising CO₂ droplets in seawater, *Environ. Sci. Technol.*, 36, 5441-5446, 2002.

Brewer, P.G., G. Malby, J.D. Pasteris, S.N. White, E.T. Peltzer, B. Wopenka, J. Freeman, and M.O. Brown, Development of a laser Raman spectrometer for deep-ocean science. *Deep-Sea Res. I*, 51, 739-753, 2004.

Caldeira, K and M. Wickett, Anthropogenic carbon and ocean pH, *Nature*, 425, 365, 2003.

Callendar, G.S., The artificial production of carbon dioxide and its influence on temperature, *Q. J. Roy. Meteorol. Soc.*, 64, 223-240, 1938.

DeLucia, E.H., J. G. Hamilton, S. L. Naidu, R.B. Thomas, J.A. Andrewes, A. Finzi, M. Lavigne, R. Matamala, J.E. Mohan, G.R. Hendrey, and W.H. Schlesinger, Net primary production of a forest ecosystem with experimental CO₂ enrichment, *Science*, 284, 1177-1179, 1999.

Dyrssen, D. and L.G. Sillen, Alkalinity and total carbonate in sea water: a plea for p-T-independent data, *Tellus*, 19b, 113-121.

Fan, S., M. Gloor., J. Mahlman, S. Pacala, J. Sarmiento, T. Takashi, and P. Tans, A large terrestrial carbon sink in North America implied by atmospheric and oceanic carbon dioxide data and models, *Science* 282, 442- 446, 1998.

Gruber, N., J.L. Sarmiento, and T.F. Stocker, An improved method for detecting anthropogenic CO₂ in the oceans, *Global Biogeochem. Cycles*, 10, 809-837, 1996.

Haugan, P. and H. Drange, Effects of CO₂ on the ocean environment, *Energy Convers. Mgmt.*, 37, 1019-1022, 1996.

Holland, E.A., and S. Brown, North American carbon sink, *Science* **283**, 1815a, 1999.

Intergovernmental Panel on Climate Change (IPCC), Climate Change: The IPCC Scientific Assessment. Cambridge University Press, 364 pp, 1990.

Intergovernmental Panel on Climate Change (IPCC), Climate Change 1995: The Science of Climate Change. Cambridge University Press, 372 pp, 1995.

Johnson, K.S., Carbon dioxide hydration and dehydration kinetics in sea water, *Limnol. Oceanogr.*, 27 (5), 849-855, 1982.

Jonsson, B., G. Karlstrom, H. Wennerstrom, S. Forsen, B. Roos, and J. Almlöf, Ab initio molecular orbital calculations on the water-carbon dioxide system. Reaction pathway for H₂O + CO₂ → H₂CO₃, *J. Am. Chem. Soc.*, 99:14, 4628-4632, 1977.

Jonsson, B., G., G. Karlsrom, and H. Wennerstrom, Ab initio molecular orbital calculations on the water-carbon dioxide system. The reaction OH⁻ + CO₂ → HCO₃⁻, *J. Am. Chem. Soc.*, 100:6, 1658-1661, 1978.

Kleinberg, R.L., C. Flaum, D.D. Griffin, P.G. Brewer, G.E. Malby, E.T. Peltzer, J.P. Yesinowski, Deep Sea NMR: Methane Hydrate Growth Habit in Porous Media and its Relationship to Hydraulic Permeability, Deposit Accumulation, and Submarine Slope Stability, *J. Geophys. Res. B*, 108, B 10, doi:10.1029/2003JB002389, 2003.

Kleypas, J.A., R. W. Buddemeier, D. Archer, J.-P. Gattuso, C. Langdon, B. N. Opdyke, Geochemical consequences of increased atmospheric carbon dioxide on coral reefs, *Science* 284, 118-120, 1999.

Langdon, C., T. Takahashi, C. Sweeney, D. Chipman, J. Goddard, F. Marubini, H. Aceves, H. Barnett, and M.J. Atkinson, Effect of calcium carbonate saturation state on the calcification rate of an experimental coral reef, *Global Biogeochem. Cycles* 14, 639-654, 2000.

Magid, E. and B.O. Turbeck, The rate of spontaneous hydration of CO₂ and the reciprocal reaction in neutral aqueous solutions between 0° and 38°, *Biochim. Biophys. Acta*, 165, 515-524.

Pasteris, J.D., B. Wopenka, J.J. Freeman, P.G. Brewer, S.N. White, E.T. Peltzer, G. Malby, Spectroscopic successes and challenges: Raman spectroscopy at 3.6km depth in the ocean. *Appl. Spec.*, 2004.

Potter, C.S., and S. Klooster, North American carbon sink, *Science* **283**, 1815a, 1999.

Riebesell, U., I. Zondervan, B. Rost, P.D. Tortell, R.E. Zeebe, and F. Morel, Reduced calcification of marine plankton in response to increased atmospheric CO₂, *Nature*, **407**, 364-367, 2000.

Shirayama, Y., Biodiversity and biological impact of ocean disposal of carbon dioxide, *Waste Mgmt.*, **17**, 381-384, 1997.

Soli, A.L. and R.H. Byrne, CO₂ system hydration and dehydration kinetics and the equilibrium CO₂/H₂CO₃ in aqueous NaCl solution, *Mar. Chem.*, **78**, 65-73, 2002.

Zeebe, R.E., D.A. Wolf-Gladrow, and H. Jansen, On the time required to establish chemical and isotopic equilibrium in the carbon dioxide system in sea water, *Mar. Chem.*, **65**, 135-153, 1999.

Zeebe, R. and D. Wolf-Gladrow, CO₂ in seawater: Equilibrium, kinetics, isotopes, *Elsevier Oceanography Series*, **65**, 346 pp, 2001.

Zondervan, I., R.E. Zeebe, B. Rost, and U. Riebesell, Decreasing marine biogenic calcification: A negative feedback on rising atmospheric pCO₂. *Global Biogeochem. Cycles*, **15**, 507-516.

Tables

Table 1. Calculated half-life of relaxation time in pH.

Temp (degree)	Depth	Relaxation time (1/2) (sec)	Memo
4.2	Lab	187	HCl injection in Lab.
9.0	“	100	“
11.7	“	87	“
13.8	“	69	“

Temp (degree)	Depth (m)	Mean Relaxation time (1/2) (sec)	Memo
10	100m	57 (n=3)	HCl injection <i>in situ</i>
6	500m	29 (n=3)	HCl injection <i>in situ</i>
6	500m	49 (n=3)	CO ₂ rich seawater injection <i>in situ</i>
4	1000m	46 (n=4)	CO ₂ rich seawater injection <i>in situ</i>

Table 2. Coefficients of the equation of relaxation time in pH.

Lab data	Fitting curve Equation	Parameters			Memo
		y ₀	a	b	
	$f=y_0+a*(1-\exp(-b*t))$	8.2418	0.1705	0.0037	HCl injection in Lab.
	“	8.0679	0.1886	0.0069	“
	“	8.0869	0.1745	0.0080	“
	“	8.0873	0.1934	0.0100	“

Depth (m)	Fitting curve Equation	Parameters			Memo
		y ₀	a	b	
100m	$f=y_0+a*(\exp(-b*t))$	8.201	0.035	0.013	HCl injection <i>in situ</i>
500m	“	7.616	0.011	0.028	“
500m	$f=y_0+a*(1-\exp(-b*t))$	7.377	0.071	0.014	CO ₂ enriched seawater injection <i>in</i>

1000
m

“

7.255

0.063

0.016

“

Figures

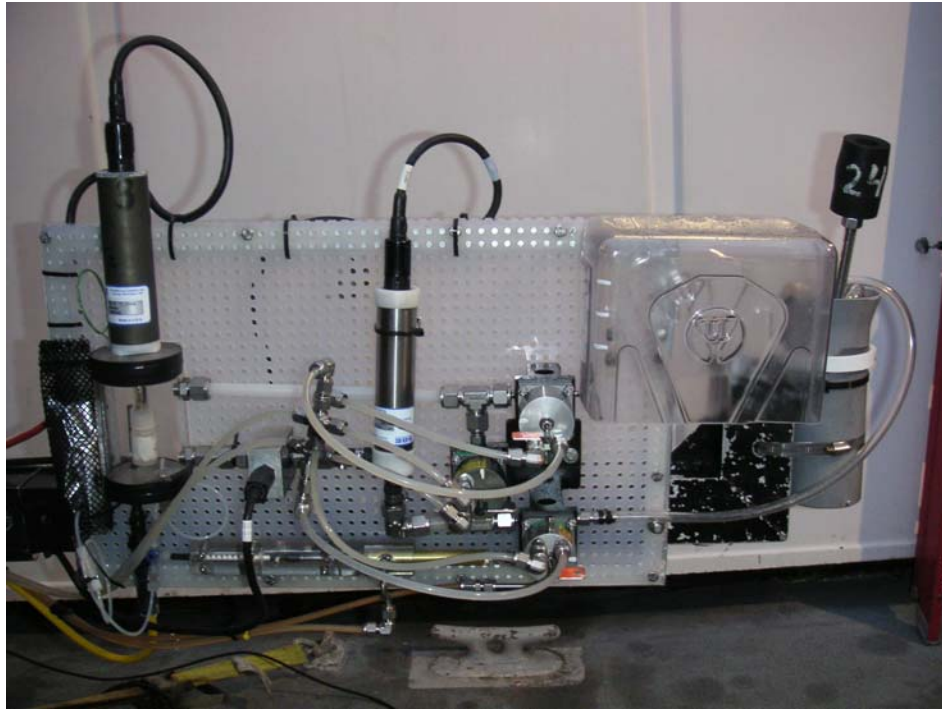


Fig. 1. A photograph of the apparatus used for the field experiment. The pH electrode is seen mounted in the titration cell at far left, in the center is the circulating pump, and just to the right of this are the two hydraulically controlled valve assemblies. At far right is the inverted clear plastic container used to hold a pool of buoyant liquid CO₂, which is delivered at depth. The handle mounted in the quiver at extreme right holds the sea water intake line, which may be positioned either to draw in normal background sea water, or CO₂ enriched water formed beneath the liquid CO₂ surface. The small plastic syringe used for acid injection into the pH cell is seen mounted horizontally at lower center; this too is actuated by hydraulic valve control from the ROV.

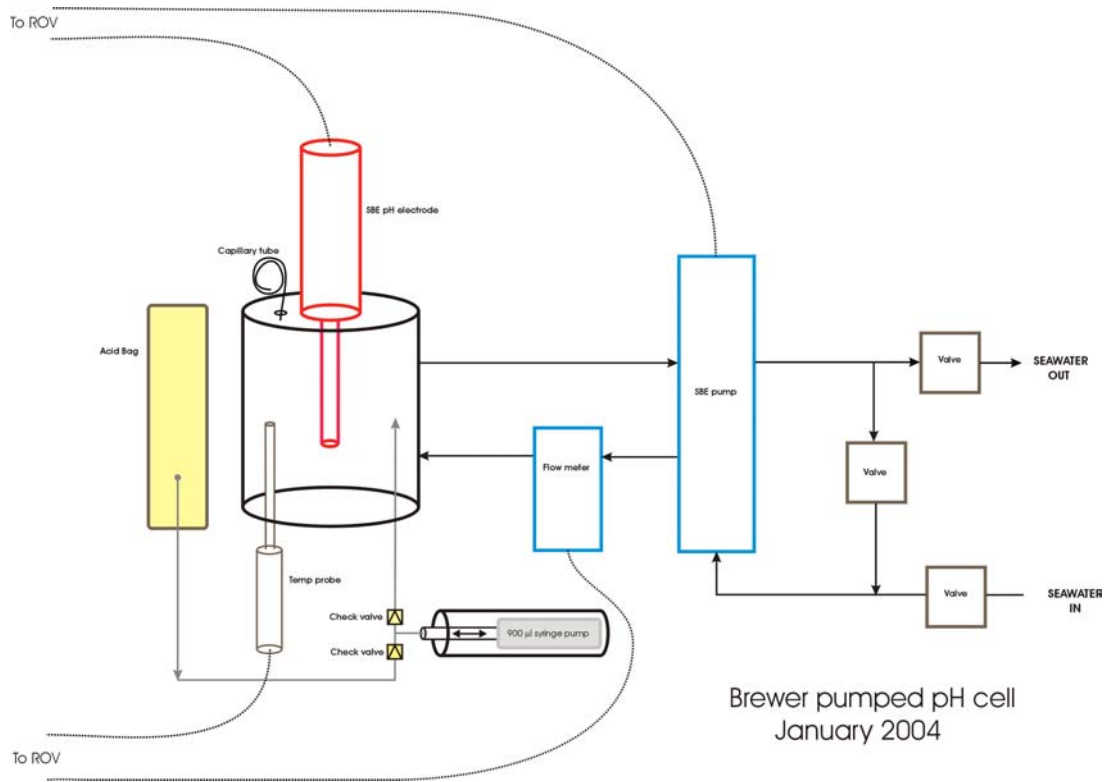


Fig. 2. Line drawing of the flow scheme for the equipment shown in Fig.1, indicating the valve arrangement and injection techniques.

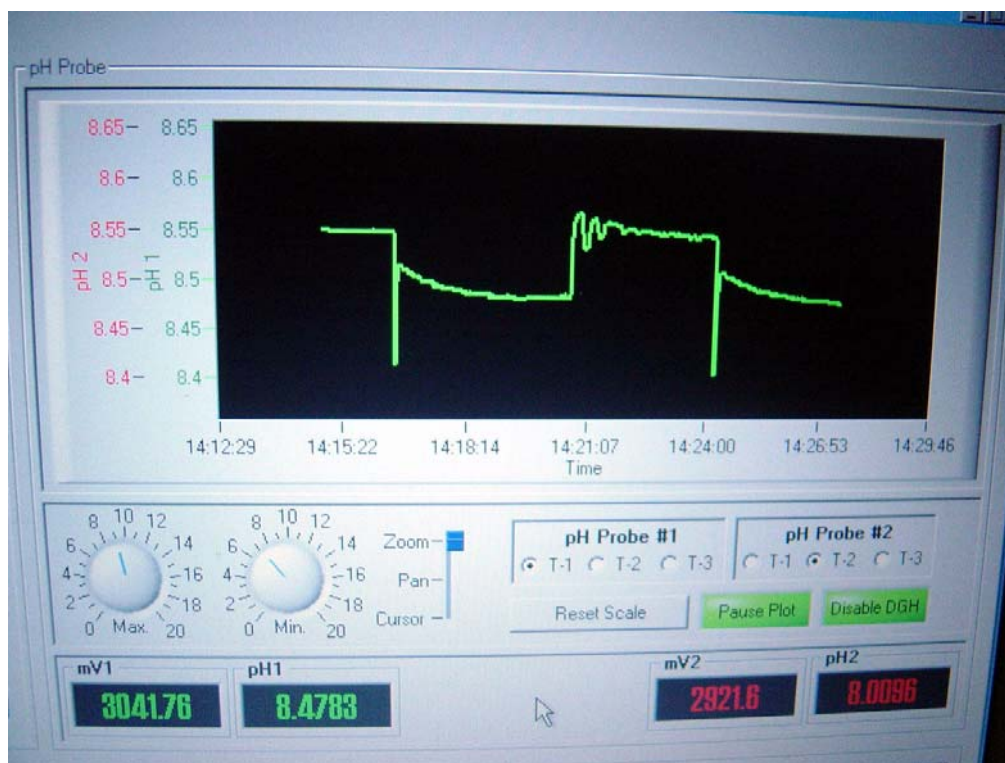
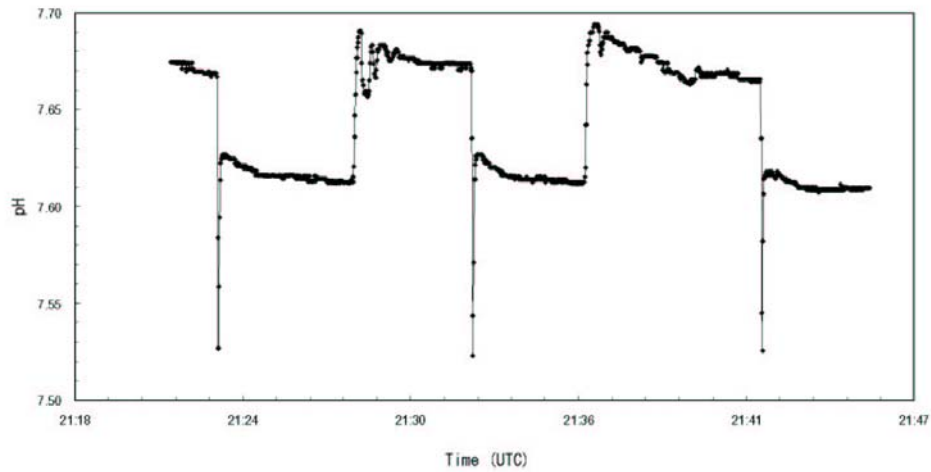


Fig. 3. Screen image of the record of injection of 900 μ L 0.008N HCl into the pH cell. Reading from left to right the trace shows a stable reading of (uncorrected) pH of \sim 8.55, followed by a sharp drop to pH \sim 8.4 as the slug of HCl is inserted. The time to stabilization at the new, lower pH is assessed from analysis of the shape of the pH-time curve observed. After 5 minutes the valves are opened several times and the system flushed. The oscillating pH signal results from venting of low pH water trapped in the dead space between the inlet and outlet valves. A second experiment is then initiated, and the sequence is repeated.

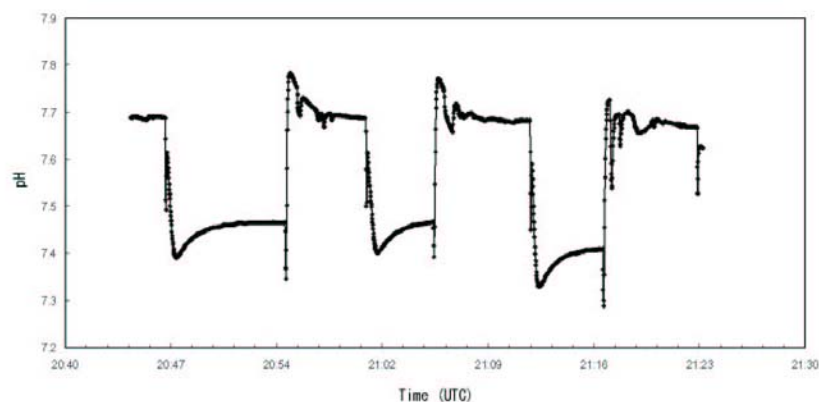
Fig. 4



D2476 Injection of HCl at 500m

Fig. 4. The data record of pH changes from a series of three injections of acid at a depth of 500m. The pH electrode output is sampled at 2 Hz. Mixing in the cell accounts for the initial rapid Δ pH, and the chemical system then relaxes towards equilibrium with a time constant defined by the shape of the pH-time curve, which here trends to lower pH with time. This is consistent with formation of an intense cloud of CO_2 at the point of injection, followed by hydration of the excess CO_2 to form HCO_3^- .

Fig. 5



D2476 Injection of CO₂ enriched seawater at 500m

Fig. 5. The equivalent data plot as for Fig. 4, but in this case a slug of sea water, highly enriched in HCO_3^- by dissolution from a liquid CO_2 interface, is drawn into the cell and re-circulated. The trend with time is to higher pH as the system adjusts to equilibrium. Both the forward and back reactions are proceeding simultaneously; the net result of a rapid decrease from injection of low pH water, followed by a slower increase in pH as the system equilibrates is consistent with removal of an excess of HCO_3^- over the equilibrium value.

Fig.A

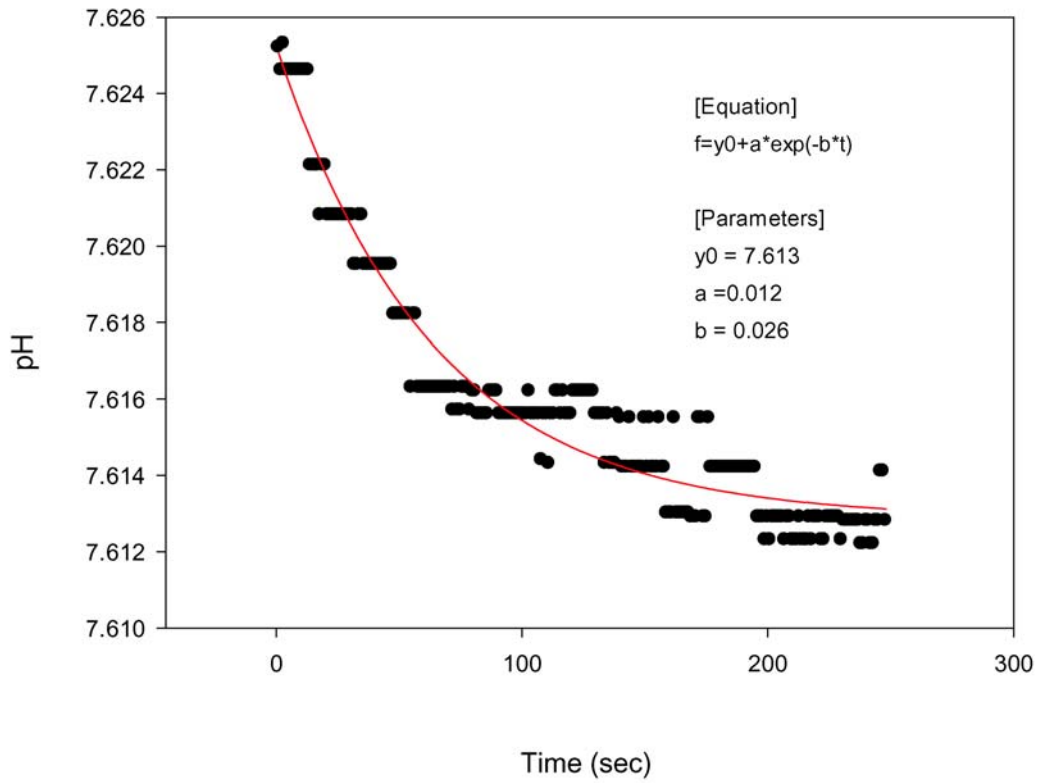


Fig. 6. Plot of pH versus time for an acid addition experiment at 500m depth, with the derived coefficients shown. The modeled data are those obtained after the initial sharp drop and rebound in pH, seen in Fig. 4 as the segment where the exponential decline to a steady value occurs.

Fig. B

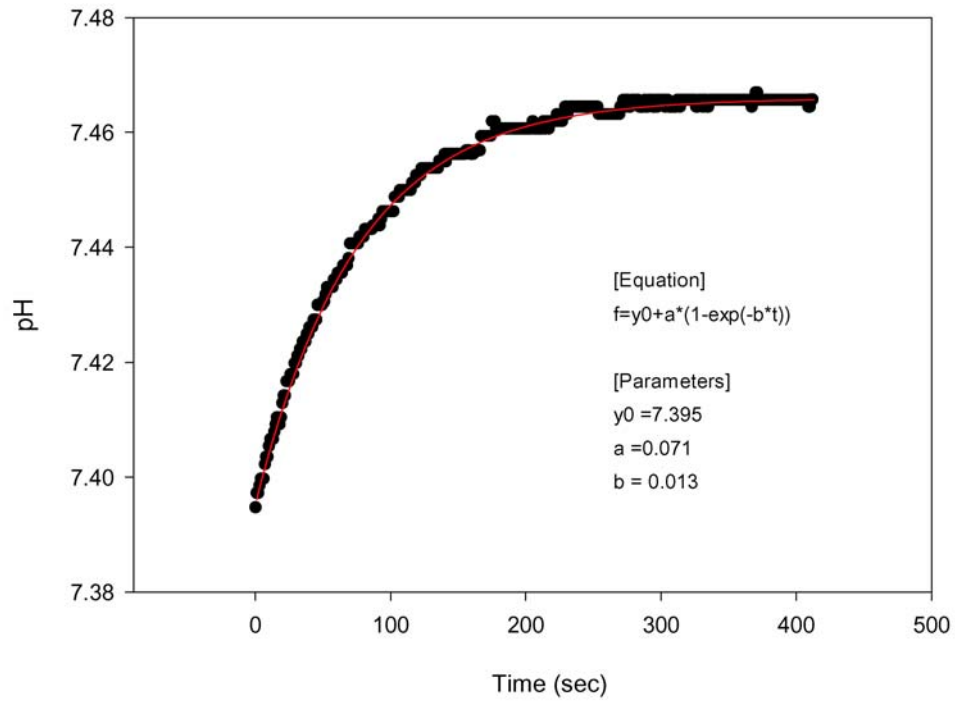


Fig. 7. The result of analysis of the relaxation time from addition of a slug of CO₂ rich water at 500m depth. Note the different trend to the acid addition experiment shown in Figure 6.

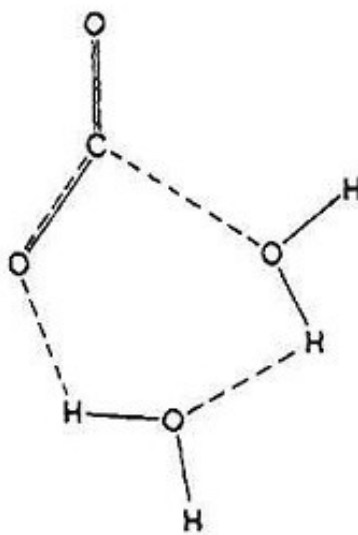


Fig 8. From Jonsson et al. (1977). Their Fig. 5 shows the transition state proposed for the reaction of CO_2 with H_2O . Their model calculations showed an unusually long C-OH distance of 1.43 Å. Since there are no barriers to reaction in the gas phase solvation effects they suggest that effects associated with the hydration sphere of the dangling proton provide a plausible explanation of the slow hydration rate observed at one atmosphere. Thus the effect of pressure on this hydration sheath also offers a plausible explanation for the higher reaction rates we report here.